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Improved Ritter reaction with CF₃-containing oxirane for an access to central units of protease inhibitors

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ABSTRACT

The influence of fluorinated alcohols on the $BF_3 \cdot Et_2O$ -promoted Ritter type reaction has been investigated. Trifluoroethanol/ $BF_3 \cdot Et_2O$ system allowed access to the central unit of protease inhibitors by the ring opening of fluorinated epoxide **2** with various nitriles.

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Diaminopropanols and corresponding amidopropanols constitute one of the most common central units of various protease inhibitors such as β - or γ -secretase,¹ HIV protease,² cathepsins,³ or plasmepsins.⁴ The scaffold **1** (Fig. 1) is a typical example of 1,3-diaminoalkan-2-ol-based structure of inhibitors.

Hydroxylethylamine motifs can be easily obtained by oxirane ring opening reactions, which furthermore allow the access of a wide structural diversity of compounds. Recently, Concellón et al. described an easy access to 1,3-diaminoalkan-2-ols by a ring opening of amino epoxides through a Ritter reaction with nitriles.⁵ This reaction permitted a direct and selective introduction of an acylamino function and offers an excellent alternative to the classical aminolysis of oxiranes. Furthermore, the reaction provided the diaminoalcohol with a protection different for each amino group, what is particularly useful for further coupling to prepare inhibitors (Scheme 1).





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In our general search for new inhibitors of aspartyl proteases,⁶ we developed an easy and diastereoselective synthesis of the trifluoromethyl epoxide **2** (Fig. 2).^{6a}

Fluorine atoms when incorporated into peptidomimetic structures or amino acids can provide interesting changes in biological properties related to altered physico chemical properties such as high hydrophobicity and higher stability toward metabolic processes.⁷ Fluoroalkyl groups are also good mimics of naturally occurring hydrophobic residues such as isopropyl or benzyl.

Ring opening of epoxide **2** can offer a general access to a wide range of polyfunctional molecules. For an easy entry to the trifluoromethyl analogue of scaffold **1**, we investigated the Ritter reaction with the amino oxirane **2**.



Figure 2.

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We first carried out the reaction with acetonitrile under reported conditions.⁸ Epoxide **2** was warmed at reflux in acetonitrile used as the solvent, in the presence of 1.2 equiv of BF₃·OEt₂. After hydrolysis, the corresponding trifluoromethyl alcohol **3a** was obtained, in a 60% yield (Scheme 2).

The ring opening of the epoxide **2** is regioselective, occurring on the terminal position (Scheme 3). For non-fluorinated epoxides, this selectivity was explained based on steric grounds. From **2**, according to the mechanism proposed by Concellón,⁸ this regioselectivity was also expected, the generation of a nitrilium ion being easier on a remote position from the electron-withdrawing CF₃ group (Scheme 3). It is interesting to note that the presence of a non-protected amine function in **2** is not an obstacle for the reaction.

The large excess of nitrile used as the solvent limits the reaction to liquids, and it is not compatible with structurally more complex nitriles. The reaction was then investigated by using CH₂Cl₂ as the solvent and nitriles as reagents under BF₃·OEt₂ catalysis (Scheme 3). With 4 equiv of MeCN, no reaction occurred at room temperature, and a complex mixture was obtained at reflux. We previously reported that hexafluoroisopropanol (HFIP) or trifluoroethanol (TFE), when used as solvents, could promote oxirane ring opening with nucleophiles.⁹ We then explored this reaction in fluorinated alcohols. Surprisingly from 2, no reaction occurred in HFIP or TFE used as solvents alone. When 1.2 equiv of BF₃·OEt₂ was added in HFIP, only traces of the desired product were obtained. This might be due to a complexation of HFIP with the nitrile function, which decreases the reactivity of the latter. The reaction was then conducted with acetonitrile in CH₂Cl₂ in the presence of BF₃·OEt₂ (1.2 equiv) and only 1.2 equiv HFIP or TFE (Table 1).

At room temperature, with 1.2 equiv of HFIP, 40% of the corresponding product **3a** was obtained after 24 h. Increasing the quantity of HFIP did not change the yield and the reaction rate. In contrast, reaction time significantly decreased when the reaction

Table 1		
Ritter reaction in DCM and ac	ctivated with fluorina	ted alcohols

Entry	MeCN (equiv)	Solvent	T (°C)	Time (h)	Yield of 3a ^a (%)
1	4	CH ₂ Cl ₂	rt	24	0
2	4	$CH_2Cl_2/HFIP(1.2)$	rt	24	40
3	4	$CH_2Cl_2/HFIP(1.2)$	40	1.5	50
4	4	CH ₂ Cl ₂ /TFE (1.2)	rt	24	67
5	4	$CH_2Cl_2/TFE(1.2)$	40	1.5	65

^a Isolated yield.

was performed at reflux (1.5 h vs 24 h). Under similar conditions, TFE appeared to be a better additive with an improved yield in product **3a** (67% at rt (entry 4) and 65% at 40 °C (entry 5)). Among different solvents used (ether, THF, ClCH₂CH₂Cl, or toluene), only CH₂Cl₂ provided clean reaction. The reaction was then extended to various nitriles under these optimized conditions (Scheme 4, Table 2).

Yields obtained in the ring opening of epoxide 2 with alkyl (entries 1 and 2), aromatic (entries 3–5), and functionalized nitriles (entries 7-9) were moderate to good, ranging from 44% to 78%. From the pNO₂ phenyl nitrile, and the reaction led to a complex mixture (entry 6), 3f could not be isolated. Clearly, the success of this reaction is due to synergic effects of trifluoroethanol and BF₃·Et₂O, since both reactions performed in TFE alone and in CH₂Cl₂ with BF₃·Et₂O were unsuccessful. This activating effect of BF₃·Et₂O in fluoro-alcohols had previously been observed in other reactions investigated in the laboratory.¹⁰ In the same connection, Prakash recently reported that BF3.2TFE complex behaves as a superacidic catalyst.¹¹ However, its preparation is not straightforward. For our Ritter reaction, we assumed that such a complex could be generated in situ and promote the ring opening. In order to validate this hypothesis, ¹H NMR experiments were performed on a mixture 1BF₃·Et₂O/2TFE in CDCl₃. In starting partners, ¹H



Scheme 4.

Table 2Ritter reaction with various nitriles



^a Isolated yield.

NMR chemical shifts of methylene protons are 3.90 ppm ($J_{H-F} = 8.5 \text{ Hz}$) for TFE and 4.17 ppm ($J_{H-H} = 7 \text{ Hz}$) for BF₃·Et₂O. In the resulting mixture, methylenes of the major compound are shielded to 4.0 ppm for CH₂-CF₃, and deshielded to 4.06 ppm for CH₂-CH₃. The shift for CH₂-CF₃ ($\Delta \delta = 0.1$) is weaker than that described for the difference between TFE and BF₃·2TFE ($\Delta \delta = 0.35$)¹¹ suggesting that the latter complex was not formed. However, exchanges between BF₃ ligands, even partial, are sufficient to promote the reaction.

In conclusion, we have described a new and easy access to trifluoromethyl diamino alcohols by the ring opening of a trifluoromethyl amino epoxide **2** by the Ritter reaction with nitriles in the presence of BF₃·OEt₂ and trifluoroethanol.¹² This method presents the great interest to provide a direct access to the central core of most protease inhibitors with a diversity of structure. The yields are moderate to good, and it is important to note that the presence of a free amino function does not influence the reaction.

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- Typical procedure for the synthesis of N-(3-benzylamino-4,4,4-trifluoro-2-12 hydroxy-butyl)-acetamide (**3a**): To a solution of trifluoromethylated epoxide **2** (100 mg, 0.43 mmol) in dichloromethane (1 mL), nitrile (1.7 mmol) and trifluoroethanol (40 µL, 0.5 mmol) were added at 0 °C. Then BF3 OEt2 (60 µL, 0.5 mmol) was introduced to the reaction mixture and warmed at reflux until the disappearance of the starting material (monitored by ¹⁹F NMR). The reaction mixture was then hydrolyzed by a saturated aqueous solution of Na_2CO_3 (5 mL), and was extracted with dichloromethane (3 \times 5 mL). The combined organic layers were washed with brine (10 mL), dried with anhydrous MgSO₄, and concentrated under reduced pressure. The crude product was then purified by chromatography on silica gel (82 mg, 65%). IR: 306; 2936; 1655 cm⁻¹, ¹⁹F NMR (188 MHz, CDCl₃, CFCl₃ as internal standard): δ –71.9 (d, *J* = 7.4 Hz, CF₃). ¹H NMR (300 MHz, CDCl₃): δ 7.24 (m, 5H), 4.64 (ddd, J = 10.1, 8.1, 2.2 Hz, 1H), 4.06 (d, J = 13.2 Hz, 1H), 3.80 (m, 2H), 3.61 (dd, J = 8.3, 13.5 Hz, 1H), 2.90 (m, 1H), 1.90 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): δ 164.5, 139.0, 128.5, 128.3, 127.5, 126.2 (q, J = 287.3 Hz, CF₃), 76.2, 59.7 (q, J = 26.4 Hz, CHCF₃), 58.0, 51.6, 13.8.